

## Kit Content

	36rxn
Syringe	36 set
Elution Tube	36 pcs
AR23100 Cartridge	36 set
AR23100 Column Set	36 set
AR23100 Tip Set	36 set
EtOH Tube	36 pcs
Sample Tube	36 pcs
Buffer DWX	25 ml
Buffer RFTL	25 ml
Buffer RFB	25 ml
Proteinase K	60 mg
Proteinase K Solvent	5 ml
Buffer RCL1	12 ml
Buffer RCL2	4 ml

## Kit Storage

Upon arrival,  
 1. Please store **AR23100 Column Set** at **4°C** for long term storage.  
 2. Please store **Proteinase K** at **-20 °C** for long term storage.  
 Buffer, solvent and consumables, please store at 15-25 °C.  
 If a precipitate has formed in Buffer RFTL, dissolve by incubating at 60°C for 10 min.

## Kit Preparation

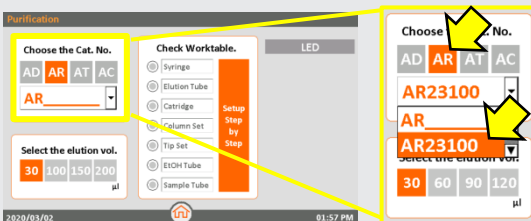
- Prepare 20 mg/ml Proteinase K**  
 For 60 mg Proteinase K, please add 3 ml Proteinase K Solvent into tube and vortex thoroughly for dissolving  
 After dissolving into solvent, please store in 4°C for 6 month or -20°C for 1 year.

## Step by Step to start a AR23100 Purification Run

- On the **Start** screen: Click “ENTER” button to enter the HOME screen.
- On the **HOME** screen: Click “Purification” icon to start a purification run.

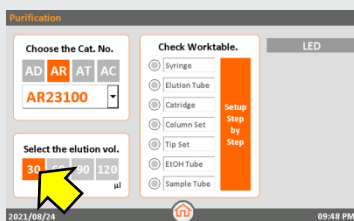


- Please choose **Cat. No.**



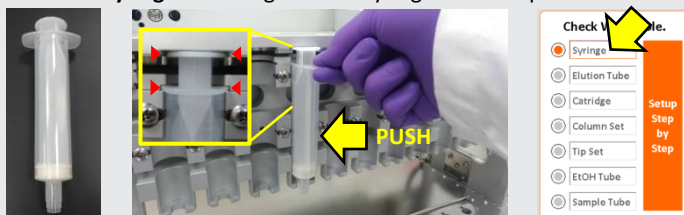
Please click “AR”  
 Then choose “AR23100”  
 For iCatcher® FFPE Tissue miRNA Kit

- Choose **Elution Vol.**



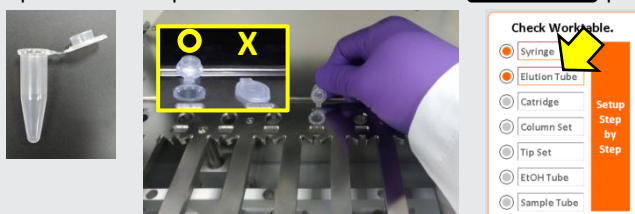
We suggest to choose **30 µl** to get higher concentration of miRNA.

- Insert the **Syringe** into the groove of Syringe Seat and push it to the end.



Check the **Syringe**.

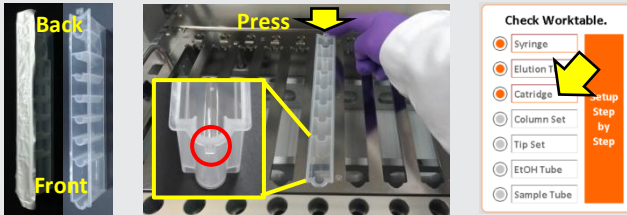
- Open the lid and place the **Elution Tube** on the **Elution Tube** position.



Check the **Elution Tube**.

**FOR RESEARCH USE ONLY**

7. Insert the front protrude part of **Cartridge** into **Cartridge** position and press the bottom down. Then remove the foil.



Check the **Cartridge**.

**Important! Please must remove the foil before running a protocol.**

8. Insert **Column Set** into **Column Set** position and press into bottom.



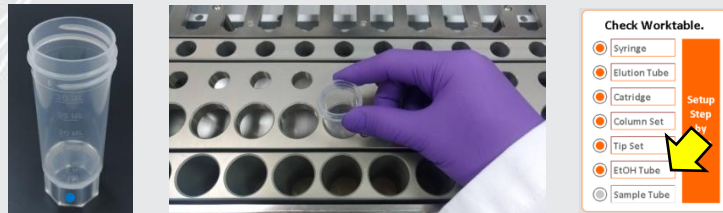
Check the **Column Set**.

9. Place **Tip Set** on **Tip Set** position and press into bottom.



Check the **Tip Set**.

10. Add **19 ml** 100% EtOH into **EtOH Tube** and place on the **EtOH Tube** position.



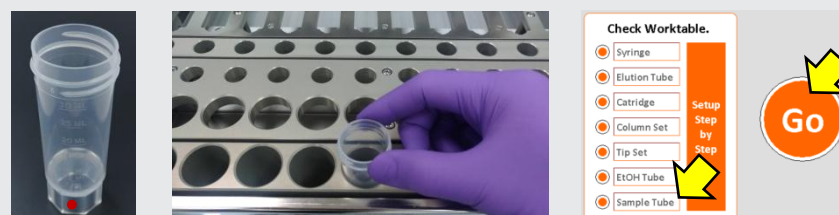
Check the **EtOH Tube**.

Add **19 ml** 100% EtOH into **EtOH Tube**

11. Prepare sample as below,

- Place 5-10  $\mu\text{m}$  sections (up to 4 sections) in a 2 ml micro-centrifuge tube. Add 600  $\mu\text{l}$  DWX buffer, vortex vigorously for 30 sec. Spin down to collect sample in the bottom.
- Incubate at 60°C for 20 min.
- Add 600  $\mu\text{l}$  Buffer RFTL (Please add 1%  $\beta$ - mercaptoethanol freshly) and mix thoroughly by vortex 10 sec.
- Centrifuge at 11,000 x g for 1 min.
- Add 80  $\mu\text{l}$  PK (20 mg/ml) to the lower clear phase. Mix gently by pipetting.
- Incubate at 60°C for 30 min. Brief spin down.
- Incubate at 80°C for 15min.
- Add 600  $\mu\text{l}$  Buffer RFB in to the lower phase, mix gently by pipetting.
- Centrifuge at 11,000 x g for 2 min. Transfer 1000  $\mu\text{l}$  lower clear phase lysate into a new 2 ml micro-centrifuge tube.
- Add 300  $\mu\text{l}$  Buffer RCL1. Pulse-vortexing for 10 sec, brief spin down then incubate at 25°C (room temperature) for 3 min.
- Add 100  $\mu\text{l}$  Buffer RCL2, pulse-vortexing for 10 sec, brief spin down then incubate at 25°C (room temperature) for 1 min.
- Centrifuge at 11,000 x g for 3 min.
- Transfer 1000  $\mu\text{l}$  clear supernatant into the Sample Tube, then add 1400  $\mu\text{l}$  Isopropanol. (no need to mix or pipette it)

12. Load the **Sample Tube** into the **Sample Tube** position of iCatcher.



Check the **Sample Tube**.

Click "**Go**" to start purification.